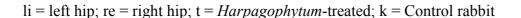
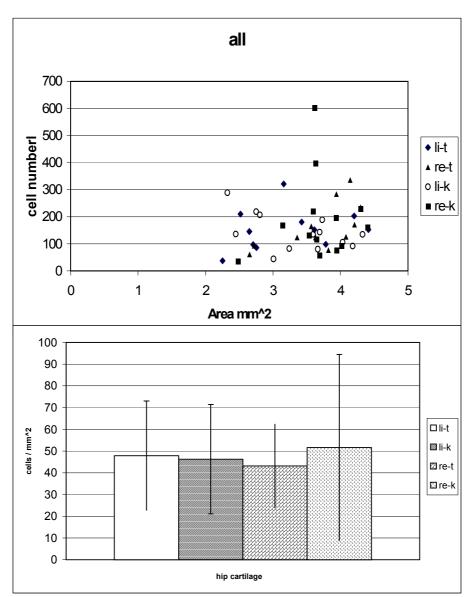
Samples were immediately immersed in 4% buffered formaldehyde and stored until further processing. For preparation of paraffin embedding, samples were dehydrated using increasing concentrations of ethanol (70%-100%) followed by immersion in xylol. After embedding in paraffin sections of 2-3µm were cut and placed on standard microscope slides. For preparation of subsequent staining, sections were dried, deparaffinized with xylol and rehydrated using decreasing concentrations of ethanol (100% - water). Sections were stained with Hemalaun for 10 minutes followed by eosin for 30 seconds. Finally, sections were dehydrated using ethanol in increasing concentrations (70%-100%), immersed in xylol and slides were mounted with Histofluid®





Cell count and area measurements on HE stainings by means of ImageJ, NIH, USA; 1.30v; URL: http://rsb.info.nih.gov/ij/

Used Macro: